Treatment of Chronic Suppurative Otitis Media With Topical Tobramycin and Dexamethasone

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Objectives: To investigate the safety and efficacy of a topical combination of tobramycin and dexamethasone in a primate model of chronic suppurative otitis media (CSOM) and to explore the contribution of the added topical steroid for the treatment of CSOM.

Design: Blinded, randomized, placebo-controlled trial.

Subjects: Sixty juvenile cynomolgus monkeys randomized into the following 6 treatment groups of 10 monkeys each: 0.3% tobramycin (group 1), combined 0.3% tobramycin–0.1% dexamethasone (group 2), combined 1.0% tobramycin–0.33% dexamethasone (group 3), 0.1% dexamethasone (group 4), vehicle (group 5), and phosphate-buffered saline solution (group 6).

Interventions: Chronic suppurative otitis media was established by inoculating the right ear with Pseudomonas aeruginosa. After 4 weeks of drainage, animals were treated according to the group assignment with 3 drops twice daily for 7 weeks. Hearing thresholds were monitored with repeated auditory brainstem response testing (ABR), and clinical response was monitored with repeated otoscopic examinations and cultures throughout the study. Cytocochleograms were evaluated for quantification of outer hair cell loss.

Results: Rapid resolution of otorrhea and eradication of P. aeruginosa occurred in all groups receiving tobramycin. The inclusion of dexamethasone accelerated the resolution of otorrhea and negative yields of cultures compared with tobramycin alone. Otorrhea and positive culture findings persisted in the groups not treated with topical antibiotic. Results of ABRs at 4 and 8 weeks and cytocochleograms for outer hair cell loss were not affected by drug administration. Perilymph samples collected at the end of the study showed no detectable tobramycin.

Conclusions: Combined tobramycin-dexamethasone ear drops were safe and effective in the monkey CSOM model. Dexamethasone enhanced the efficacy of tobramycin.


TREATMENT of otorrhea through a perforation or ventilation tube is a challenge for clinicians. Often, common respiratory pathogens are isolated.1 However, unlike otitis media behind an intact tympanic membrane (TM), pathogens such as Pseudomonas aeruginosa, Staphylococcus aureus, and other gram-negative organisms commonly are found, likely gaining access to the middle ear (ME) via the external auditory canal.2 Many commonly prescribed oral antibiotics are not effective against these more troublesome bacteria in otorrhea.3,4 Ear and eye drops containing an antimicrobial agent, with or without an anti-inflammatory component, have commonly been used for the treatment of acute or chronic otorrhea. A survey of pediatricians revealed that 79% of the respondents prescribed topical medication for treatment of chronic suppurative otitis media (CSOM).3 Another survey revealed that 84% of the otolaryngologists use ototopical preparations for otorrhea through a perforation, 94% in the presence of a ventilation tube, and 93% in a draining tympanomastoid cavity.6 Eighty percent of otolaryngologists believed that the risk of sensorineural hearing loss (SNHL) due to infection was as great as or greater than that due to the topical agents. Despite this belief, 3.4% of the same respondents claimed to have observed ototoxic effects related to ototopicals.6

The ototoxic effects of various systemically applied drugs are well documented in the literature.7 Whether topical aminoglycosides applied to an inflamed human ME are ototoxic remains controversial. Most of the data in support of ototoxic effects in humans are anecdotal case reports.5,8 Only a few studies have looked
MATERIALS AND METHODS

Sixty juvenile cynomolgus monkeys of both sexes were included in the study. The animals were quarantined for 4 weeks before the experiment. All animals were examined using an operating microscope and tympanometry to confirm the health of the TM and ME. The animals were assigned randomly (10 animals per group) to 1 of the following 6 treatment groups: group 1, 0.3% tobramycin; group 2, 0.3% tobramycin–0.1% dexamethasone; group 3, high-dose 1.0% tobramycin–0.33% dexamethasone; group 4, 0.1% dexamethasone; group 5, vehicle containing benzalkonium chloride; and group 6, phosphate-buffered saline (PBS) solution. This inoculation was repeated a week following 6 treatment groups: group 1, 0.3% tobramycin; group 2, 0.3% tobramycin–0.1% dexamethasone; group 3, high-dose 1.0% tobramycin–0.33% dexamethasone; group 4, 0.1% dexamethasone; group 5, vehicle containing benzalkonium chloride; and group 6, phosphate-buffered saline (PBS) solution. Histological findings and auditory brainstem response (ABR) data on the uninfected, untreated left ears were collected and are presented as separate control data for comparison of normative data. Investigators and study personnel were blinded to the treatment group assignment. The procedures on animals were approved by the Animal Research and Care Committee at the Children's Hospital of Pittsburgh, Pittsburgh, Pa, and in accordance with the US Public Health Service Policy on Humane Care and Use of Laboratory Animals, the National Institutes of Health Guide for the Care and Use of Laboratory Animals, and the Animal Welfare Act.

STUDY DESIGN

This study enabled investigation of the efficacy and safety of tobramycin and dexamethasone at 2 different concentrations, as well as the components 0.3% tobramycin, 0.1% dexamethasone, and vehicle as separate treatments for the assessment of effects of each component on the outcome. A sixth group included as a negative control group was treated with PBS drops. Noninfected, untreated, and untreated left ears of 2 animals from each of the 6 treatment groups were preselected randomly for comparison with the treated ears. The ABRs were performed only once on the left ears in this group. Middle ear and perilymph samples were obtained and the cochleae were processed to assess ototoxic effects and to compare normative data for the intact ears.

A baseline ABR was performed to document the hearing thresholds of all right ears, after which a wide perforation of the TM (approximately 75%) was created. This was followed by a second ABR. The right ears then were inoculated with 10^6 colony forming units of P aeruginosa (Rochester strain). This inoculation was repeated a week later in all animals regardless of their otorrhea or culture status. The animals were observed without any treatment for 4 weeks. During this period, the ears were examined twice a week under the operating microscope, cultures were obtained without suctioning, and findings were recorded. At the end of 4 weeks of drainage, a third ABR was performed on all right ears. Then topical treatment was begun according to group assignment. The medications were delivered from uniform, color-coded droppers, blinding the study personnel and examiners to group assignment. The pH of the ear drops was 7.5, 5.5, 5.6, 6.6, 5.67, and 6.02 for the groups 1, 2, 3, 4, 5, and 6, respectively. Twice daily for 7 weeks, 3 drops were placed into the right ears. The animals were examined otomicroscopically twice a week, a culture was obtained, and the external ear canals were suctioned. A fourth ABR was performed on all right ears after the fourth week of treatment. The treatment was discontinued after 7 weeks, with continued otoscopic and culture evaluations until the animals were killed. A fifth ABR was performed 1 week after discontinuation of the treatment. All the animals were killed approximately 1 week after the last ABR. On the day the animals were killed, a drug penetration study was performed on selected animals to determine the amount of tobramycin in the perilymph. The animals were killed under heavy anesthesia using vital perfusion with a fixative. The temporal bones were dissected, gross macroscopic findings on the TM and ME were recorded, and biopsy specimens were obtained. The cochleae were perfused with fixative in situ through the round and oval windows and then dissected from the rest of the temporal bone and prepared for cytocochleogram.

SPECIFIC METHODS

Ketamine hydrochloride (10 mg/kg) was used for the anesthesia of the animals for brief procedures such as the otoscopic examinations with culturing and ear drop application. For the ABR, the animals were anesthetized with a mixture of ketamine hydrochloride (13.3 mg/kg), xylazine hydrochloride (2.7 mg/kg), and acepromazine maleate (0.4 mg/kg). The supplement was given as needed using half the initial dose.

Otoscopic examinations were performed with the operating microscope (M703F; Storz Instrument Co, St. Louis, Mo) at 16× magnification. Sterile neonatal speculi (Storz Instrument Co) and pediatric disposable speculi (Kleen-spec; Welch Allyn, Skaneateles Falls, NY) were used during otomicroscopy. The otoscopic findings were recorded for presence of perforation (perforated, not perforated, or could not evaluate), presence of drainage (yes or no), quantity of drainage (0 indicates no drainage; 1, moist; 2, filling the ME; 3, filling the external auditory canal; and 4, draining out from the external auditory canal), and quality of drainage (serous, mucoid, mucopurulent, or purulent).

A culture was obtained during the otoscopic examination using a calcium alginate fiber–tipped aluminum applicator swab (Fisherbrand Sterile Swabs; Curtin Matheson Scientific, Houston, Tex). The swab was streaked immediately on chocolate agar plates (for isolation of any bacteria present) and Pseudomonas isolation agar (Difco Laboratories, Detroit, Mich) plates (for P aeruginosa). The plates were placed in an incubator at 37°C and read at 24, 48, and 72 hours.

After 4 weeks without treatment, the ears were suctioned (sterile 5F or 3F cannulas; Storz Instrument Co) after at ototoxic effects in humans in a systematic way.10-12 The small number of studies that relate topical treatment to hearing loss in humans probably results from the difficulty in differentiating the effects of coexisting chronic or recurrent infection from that of topical aminoglycosides, since both potentially affect hearing.9,10

Until recently, no ototopical antimicrobial preparation had been approved by the US Food and Drug Administration (FDA) for treatment of otorrhea in the face of a nonintact TM. Clinicians managing chronic and recurrent otorrhea have used such preparations without an FDA-approved otic indication, after warning the pa-
samples were obtained for cultures. The presence of the perforation was checked, and if the TM had healed, it was reperforated. If the size of the perforation was smaller than 50% of the TM area, it was enlarged. Reperforation of the TM was documented.

The ABRs were performed in a soundproof booth using a compact auditory electrodagnostic system (Nicolet Instrument Corp, Madison, Wis) for stimulus generation and potential recordings. Two types of stimuli were presented: clicks and tone bursts of 2000, 4000, and 8000 Hz. For each stimulus, 2000 sweeps were recorded, analyzed, and stored. Stimulus intensity was decreased from 100 dB, in steps of 20 dB down to 40 dB, in steps of 10 dB down to 20 dB, and in steps of 5 dB down to the level of hearing threshold. When ABR potentials disappeared, the ABR was repeated at 5-dB increments until the potential disappeared. The level of hearing was then confirmed by obtaining the potential again at the 5-dB lower threshold.

After the sedation with ketamine hydrochloride (10 mg/kg), animals were killed using intraperitoneal injection of pentobarbital sodium (35 mg/kg), followed by vital perfusion an hour later. Intracardiac perfusion with PBS for 3 minutes was followed by perfusion with Karnovsky fixation solution for 10 minutes. The temporal bones were dissected out within half an hour. The bony external ear canal was removed, and the TM thickness was graded (scale, 0–4+) using an operating microscope. Following removal of TM and malleus, the thickness of ME mucosa (scale, 0–4+) was assessed. Thicknesses were graded by the same investigator (C.M.A.), to have a subjective assessment of the differences between specimens. A punch biopsy specimen, including bone periosteum and mucosa, was obtained from the hypotympanum. The incus and stapes were removed and saved for histological examination. A separate bone and mucoperiosteal punch biopsy specimen was obtained from the mastoid, just peripheral to the antrum. The tissues were decalcified with 5% formic acid, embedded in paraffin, sectioned, and stained with hematoxylin and eosin using standard techniques. The thickness of the TM and the mucosa of ME and mastoid were measured as an indicator of the degree of inflammation, using an image analysis package. Quantitative histological analysis was performed using a commercially available software (Metamorph Imaging System, Version 2; Universal Imaging Corporation, West Chester, Pa). Mucosal thickness was measured on the sections stained with hematoxylin and eosin. For each location, 3 measurements were randomly obtained, and their average was used in the analysis.

Immediately after the luxation of the stapes, the perilymph was aspirated through the oval window with a microliter pipette (Pipetman; Rainin Instrument Co, Woburn, Mass) and saved. The round window was punctured, and the labyrinth was perfused with fixative. Following trimming to remove the bone around the labyrinth, the cochlea was placed in the fixative in a refrigerator at 4°C overnight. The cochlea were placed into cacydolyte buffer and reperfused through the round window twice weekly until shipment to the Karolinska Institute, Stockholm, Sweden. There, the specimens were dehydrated and embedded in agar. They were processed in a routine manner for light microscopy and dissected for cytocochleogram.

On the day each animal was killed, the treatment code was broken and a perilymph penetration study was performed. The groups that had received drops with tobramycin during the treatment period were used for the study. The left ears of a few animals were included as controls. According to the protocol, the treatment had been discontinued at least 1 week before the animals were killed. To assess the penetration of tobramycin through the round window membrane, 3 drops of 0.3% tobramycin were applied in vivo to the external ear canals, similar to the method used throughout the experiment. To detect any potential time dependence, the animals were divided into 2 groups, and perilymph samples were collected approximately 1 or 3 hours after the dosing. After the animals were killed and the temporal bone was dissected, the ME was irrigated thoroughly to wash out any remaining tobramycin. The ME was dried, and the stapes was luxated. A micropipette with a pipet tip was used to aspirate the perilymph. The sample volume was measured, and it was stored at 80°C until the transfer and the analysis for the assay. To control for the effect of perilymph contamination from the ME during sampling, a group of left ears was dissected as described, and 3 drops of tobramycin were applied. The ME was irrigated and dried as in the experimental group just before obtaining the perilymph sample.

The perilymph samples were transferred to TexMS Analytical Services, Houston, Tex, for quantitative analysis of tobramycin. An internal standard was prepared with bekanamycin in perilymph fluid. Aliquots were analyzed by means of high-pressure liquid chromatography and mass spectrometry with electrospray ionization and selected ion monitoring. The protonated molecular ions for tobramycin (mass to charge ratio, 490) and the internal standard bekanamycin (mass to charge ratio, 506) were monitored, and the resulting intensity ratio was used for quantitation. A 5-point calibration curve was then generated using control perilymph matrix spiked with tobramycin at 40, 60, 80, 100, and 120 pg/µL. Duplicate calibration standards were used at the high and low ends of the curve. Single standards were used for the 3 intermediate levels. Separately prepared perilymph samples spiked at 60 and 100 pg/µL were used as quality control samples and analyzed in duplicate.

A block-surface technique method described by Spoendlin and Brun13 was used for the histological evaluation of ototoxic effects. Surface preparations were examined under light microscopy for the quantification of hair cell loss. The total number of outer hair cells (OHCs) and of damaged OHCs were counted in 4 parts (each coil in 2 parts). Hair cells were counted separately for each position (first, second, and third OHC rows) in each part of each coil. The results are reported as percentage of the damaged hair cells for each part of the cochlea.

A combination of tobramycin and dexamethasone is currently marketed as an eye drop; however, it has been prescribed without an FDA-approved otic indication by otolaryngologists for the treatment of otitismedia as well. We therefore conducted a study to investigate the efficacy and safety of topical tobramycin-dexamethasone in
Pretreatment and posttreatment periods, is illustrated in Figure 1. The drainage was present in more than 95% of the observations across all groups, before initiation of treatment. All study groups had an average of grade 3 corresponding to drainage at least filling the external ear canal. The amount of drainage remained constant for groups 4, 5, and 6 throughout the treatment period. However, there was a gradual decrease in drainage for the groups treated with drops that included tobramycin (groups 1, 2, and 3). In groups 2 and 3, the average score for drainage decreased in just 2 weeks to 2, corresponding to filling only the ME, and in 4 weeks to 1, corresponding to moisture in the ME. The rate of decrease in the amount of drainage was faster for groups 2 and 3 when compared with group 1. The average score for otorrhea in group 2 decreased to less than 1 by the fifth week of treatment and remained stable at that level for the remainder of the study. On the other hand, group 1 had a lower rate of decrease in the amount of drainage. Furthermore, the drainage increased in group 1 when the ear drops were discontinued at the end of the study.

**CULTURE FOR P AERUGINOSA**

*Pseudomonas aeruginosa* was isolated from ear cultures in 98% of the observations before initiation of the treatment. There were no differences in culture positivity for *P aeruginosa* between groups during the pretreatment period. The cultures continued to yield positive results throughout the treatment period for *P aeruginosa* for groups 4, 5, and 6 (Figure 2). On the other hand, the groups that received drops containing tobramycin, ie, groups 1, 2, and 3, had a rapid decrease in the percentage of ears with cultures yielding *P aeruginosa*. All of the ears in group 2 yielded negative culture findings after 3 weeks of treatment, and these remained negative for the rest of the study period. In group 3, the ears with culture-positive findings decreased rapidly to 20% in 1.5 weeks; however, complete eradication did not occur until another 4 weeks of treatment had passed. Group 1 had a gradual decrease across 6 weeks to reach 0% *P aeruginosa* in cultures. The
generally continued to yield negative findings during the last week of the study, when ear drops were not administered. However, a relapse for *P. aeruginosa* during the ninth week was observed in 2 of 6 ears in the interval between the end of treatment and killing.

### ABR TESTS

The hearing thresholds of the right ears underwent ABR testing 5 times during the study. At entry, the right ears had average (± SD) hearing thresholds of 21.4 ± 7.4, 27.6 ± 7.6, 30.8 ± 10.8, and 34.5 ± 12.5 dB for click and 2000-, 4000-, and 8000-Hz stimuli, respectively. Perforating the TMs resulted in 29.0-, 31.8-, 25.9-, and 24.3-dB increases in the thresholds for click and 2000-, 4000-, and 8000-Hz stimuli, respectively. Perforation between the groups. The average thresholds for each treatment group at baseline, postperforation, pretreatment, treatment-interval, and posttreatment ABRs are shown in Table 1. The uninfected, untreated left ears served as controls, with an average score of 0 for TM and ME. Groups 3, 4, and 6 had higher scores for inflammation. Group 1 had the lowest score among the infected study groups.

### ME FINDINGS

Average scores for the degree of TM and ME inflammation determined by otomicroscopy are presented in Table 2. The uninfected, untreated left ears served as controls, with an average score of 0 for TM and ME. Groups 3, 4, and 6 had higher scores for inflammation. Group 1 had the lowest score among the infected study groups.

### COCHLEAR HISTOPATHOLOGICAL FINDINGS

The light microscopic evaluation demonstrated purulent material within 2 cochleae, 1 in groups 5 and 6 each. In those specimens, the inner ear structures were grossly normal.
identified; however, damage to the organ of Corti and significant loss of hair cells prohibited any count from being performed. The data of the 2 cochleae with pus are not included in the calculation of the average OHC loss (OHCL) of these treatment groups. The cytocochleogram revealed partial damage, probably as a result of processing, in 2 other cochleae. The OHCs in the first coil of a cochlea in group 6 and all the OHCs except the first part of the first coil in another cochlea in group 5 could not be read. Only the data on undamaged sections of these 2 cochleae are included in the calculation of the averages of the treatment groups.

Results of light microscopy demonstrated normal basilar and Reissner membranes, stria vascularis, and organ of Corti (Figure 4, A). There was no damage to the inner hair cells (Figure 4, B and C). The OHCL did not show considerable differences between the various parts of cochlea (Table 3). The OHCL in the first coil was slightly higher than in the second coil, a finding consistent throughout the groups, including the control group. The average OHCL was 0.90%, 0.86%, 1.11%, 1.09%, 0.90%, 0.82%, and 0.65% of the total number of hair cells for groups 1, 2, 3, 4, 5, and 6 and the control group, respectively.

None of the cochleae in any treatment group, except the 2 with pus, had more than 3% OHCL. Although OHCL below the levels of 5% is considered insignificant, the distribution of the number of ears that had at least 1% and 2% OHCL in each treatment group are presented in Table 3. These results suggest no significant OHCL during the topical treatment of CSOM with tobramycin or dexamethasone.

### COMMENT

The ethical and methodological limitations of obtaining reliable information on otoxic effects in humans leads to research in animals. Most of the studies on ototoxic effects have been performed in chinchilla or guinea pig models. Several studies in various animals have demonstrated hearing loss or cochlear damage due to ear drops. Various antibiotics, antifungals, and solvents penetrated the perilymph and produced hair cell loss in animals. When the same ear drops were placed into the ME of higher species, such as baboons, a more limited sensory cell loss was seen. On the other hand, topical dosing of ciprofloxacin hydrochloride to chinchillas demonstrated no significant effect on ABRs or the cytocochleogram. The outcome differences in these studies may result from active ingredients in the drops, duration of the treatment, degree of inflammation, and differences in size, thickness, location, and permeability of the round window membrane between species. Applicability of these safety results to humans, therefore, have been questioned.

The differences in human ME architecture has implications not only for safety but also for treatment efficacy. Studies on ototoxicity are rarely conducted in an infectious model. This results in part from the lack of a good infectious model in rodents. Significant anatomic interspecies differences in the external ear canal, eustachian tube, mastoid pneumatization, and access to mastoid from ME have limited the use of animals for efficacy studies. Mastoid involvement in human CSOM has been claimed to be a major factor in the chronicity. The restricted access between the ME and mastoid air cells was thought to limit the efficacy of ear drops. These concerns, besides the concerns about applicability of rodent safety results, support basic studies in primates. Therefore, a monkey animal model was developed by Dohar et al to study efficacy and safety of ear drops in CSOM.

The primary reason for preferring a noninfectious model for evaluating safety is to test the ototoxicity of an ear drop when the ear is in its most vulnerable state. Although ear drops are typically prescribed to treat drainage, drops may be used after the resolution of drainage. The persistence of inflammation in the ME may continue to limit the penetration of drops into the perilymph. The undetectable levels of tobramycin in the perilymph in our study may reflect restricted permeability due to residual ME inflammation, despite presumed cure seen on otoscopy and culture findings. In our study, the degree of inflammation as assessed by mucosal thickness in the groups with persistent infection was up to 8 times that of the control side for the TM, 5 times that for the ME mucosa, and 3.5 times that for the mastoid mucosa. The degree of ME inflammation in our study appeared to be mainly due to infection. However, although otorrhea had resolved before the end of the study in the groups treated with tobramycin-containing drops, inflammation (mucosal thickness) persisted in TM, ME, and mastoid mucosal specimens. Even in the group with the least degree of inflammation, group 2, TM thickness was 4 times and ME mucosa thickness was 1.5 times the control side. The thick-
ness of the mastoid mucosa in group 2 was the same as that of the control group. Group 1, however, had relatively higher average thickness at the time the animals were killed. The only group with recurrent *P. aeruginosa* infection (group 1) had a similar amount of mastoid inflammation as the groups with persistent drainage. This is reflected in the thicker mastoid mucosa. On the other hand, some investigators linked the biofilm state of the bacteria with the persistence or recurrence in CSOM and other chronic infectious diseases. These bacteria are distinct from their planktonic forms, very resistant to antibiotics and host defense mechanisms, and difficult to isolate using routine culture techniques. However, it is unclear, given this explanation, why only a few ears in group 1 had recurrence in our study.

In a study on susceptibility patterns of aural *Pseudomonas* isolates, Dohar et al reported that tobramycin had significantly better in vitro activity (94%) compared with gentamicin (79%). Piperacillin sodium was only intravenous agent with better (96%) in vitro activity. Our study clearly demonstrates the efficacy of tobramycin in the resolution of otorrhea and elimination of *P. aeruginosa*. All 3 groups treated with tobramycin (groups 1, 2, and 3) had gradual resolution in the amount of otorrhea over several weeks, whereas there were no differences in the amount of otorrhea in the other 3 groups (groups 4, 5, and 6). Both groups receiving tobramycin-dexamethasone (groups 2 and 3) experienced more rapid resolution of otorrhea than did the group receiving tobramycin alone (group 1). These results may in part be consistent with the study by Fradis et al that compared the efficacy of topical tobramycin and ciprofloxacin for the treatment of CSOM in humans. In their study, tobramycin (without dexamethasone) was found to be equally effective (66.7%) in bacteriologic response but worse in clinical response (72.2% vs 78.9%) when compared with ciprofloxacin ear drops. In another study, ototopical ciprofloxacin was found to be effective in nearly 70% of patients with otorrhea associated with *P. aeruginosa*, previously unresponsive to other antimicrobials.

In a study of children with acute purulent otorrhea, topical ofloxacin demonstrated an 84.4% cure in subjects evaluated clinically, and it eradicated 96.3% of all baseline pathogens in subjects evaluated microbiologically. Our study clearly demonstrates an added benefit of dexamethasone when combined with tobramycin for resolving otorrhea and eradicating *P. aeruginosa*. The duration of treatment necessary to achieve complete eradication of *P. aeruginosa* from the ears was 3, 5, 5, and 6 weeks for groups 2, 3, and 1, respectively. Although not quite different than the 66.7% bacteriologic response reported with 3 weeks of topical treatment of human CSOM with ciprofloxacin or tobramycin in the study by Fradis et al, prolonged need for treatment in the monkey model may result from less frequent (twice a day) administration of the drops, limited access of the dropped medications into the ME because of the narrow ear canals, and anticipated natural nonhygienic habits of monkeys that may lead to constant or recurrent contamination of their perforated ears.

Differences in the responses for culture and clinical outcome measures were observed in a previous study using this model. In that study, a rapid eradication of *P. aeruginosa* with topical ciprofloxacin, its vehicle, or Cortisporin (a combination product of hydrocortisone, neomycin sulfate, and polymyxin B sulfate) was not followed by a clinical response as assessed by otoscopy. Although the treatment period in that study was only 4 weeks, no decrease in the amount of otorrhea was observed. However, in our study, a response was apparent even in the first 4 weeks of the 7-week treatment period. In fact, for groups 2 and 3, the drainage score was halved in the first 2 weeks.

Monitoring of hearing loss is conducted when medications with potential ototoxicity are used in clinical practice or in animal studies. Hearing loss was monitored with ABR at 5 different stages throughout our study, to better identify, besides the histological features, the contribution of each of the potential factors, ie, TM perforation.
Table 3. Inner Ear Findings in 6 Treatment Groups and Controls*

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<th>No. of cochleae with loss</th>
<th>1</th>
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<td>1</td>
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<td>0</td>
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<tr>
<td>Partially damaged</td>
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*Groups are described in the second footnote to Table 1. Controls consisted of intact left ears. †Data are given as average ± SD.

otitis media with effusion.28 When effective medications are used, this complication of CSOM is rarely seen. This significant complication in our study reminds us of all the potential complications of CSOM, some of which are life threatening, and the importance of developing a safe and effective treatment for this condition.

The data on the OHCL did not include the 2 ears infected with Pseudomonas aeruginosa, but before the initiation of the treatment. The analysis of hearing loss and histological results for each individual animal with relevant data did not demonstrate an ototoxic effect of tobramycin or dexamethasone.

A Pearson correlation analysis between OHCL and change in hearing with treatment in all of the animals did not demonstrate a statistically significant ($r = -0.12; P = .39$) relationship. It should be remembered that the data for cochleae with pus or with damaged sections were excluded from the initial analysis. To assess the effect of this on the
correlation, 100% of OHCL was entered to the sections of missing data because of pus or damage. This brought the correlation coefficient to 0.33 at \( P = .01 \). It seems that the persistence of otorrhea and the chronicity of the supplicative otitis media may increase the OHCL. The SNHL in some animals seems to be due to the infection itself. At least in the 2 cochleae with pus, this relationship is clear. The correlation between hearing and histological findings was analyzed separately only for the 3 groups that were given tobramycin. This correlation between hearing and OHCL for the groups was significant (\( P = .03 \)) for groups 1, 2, and 3. However, the correlation for these groups was negative (\( r = -0.40 \)), suggesting that for groups that received tobramycin, more pronounced hearing loss was associated with less OHCL. This negative correlation may also mean that with the resolution of otorrhea, there was an improvement in hearing, but that this may be increasing the risk for OHCL. Although the degree of OHCL in this study was well below the thresholds for ototoxic effects, this slight trend is consistent with other studies that have demonstrated the effect of inflammation in reducing the risk for ototoxic effects.

Our study monitored otoxicity, similar to other animal studies, by assessing the hearing loss and OHCL. This approach ignores potential vestibulotoxicity. No apparent signs of vestibulotoxic effects developed in any of the animals during the study period; however, a systematic and objective method of assessment was not part of the study protocol. Since many topical agents, such as aminoglycosides, may have prominent toxic effects on the vestibule, some while sparing the cochlea, standard test batteries should be developed to monitor vestibulotoxicity and included in the animal models for ototoxic effects.

Our study suggests that tobramycin with dexamethasone is a very effective treatment for otorrhea and eradication of \( P \) aeruginosa in a monkey CSOM model. When comparing the treatment groups for hearing threshold changes and OHCL, our study suggests that neither tobramycin nor dexamethasone is ototoxic, individually or in a combined formulation. Dexamethasone clearly contributes to the efficacy of the combination ear drop, leading to more rapid resolution of the CSOM and eliminating the risk of recurrence after the discontinuation of drops. The tobramycin-dexamethasone ear drop presents a promising choice for the treatment of otorrhea due to \( P \) aeruginosa infection. Future studies should address potential vestibulotoxicity of aminoglycosides as well as assess the safety and efficacy of this ear drop in humans.

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